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Kona-line SPF shrimp, Vibrio penaeicida candidates for bacterial challenge model

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Direct relationship observed between bacterial concentrations and shrimp mortality

Most shrimp disease challenge models have been developed and standardized for the study of diseases caused by such specific viruses as White Spot Syndrome Virus and Taura Syndrome Virus. These challenge models are an important component of shrimp-breeding programs as an aid in disease-resistance selection. They are also used to evaluate the effects of environmental factors on the manifestation of the diseases and in the evaluation of possible therapeutic and preventative agents.

Bacterial challenge models, in comparison, are less developed. Although very important, the economic impacts of bacterial diseases on the shrimp-farming industry are apparently not as severe as those of some viral diseases, so less interest and research funding are channeled in that direction.

Additionally, bacterial challenge models are very difficult to standardize because shrimp's responses to infection are highly dependent on their physiological condition, the source of the shrimp, bacterial species and strains, route of infection, environmental conditions, and other factors. Moreover, some of the most common bacteria pathogenic to shrimp – usually *Vibrio* species – are opportunistic and the cause of secondary infections.

For challenge studies where healthy shrimp are employed, unusually high numbers of these bacteria are necessary to cause disease. However, such high numbers of bacteria also tend to quickly overwhelm the shrimp's immune systems, and the shrimp succumb to the disease before any additional testing of therapeutics can be done.



The experimental units consisted of covered 90-liter plastic tanks fitted with internal biological filters.

Challenge model

The Kona-line shrimp – specific pathogen-free (SPF) *Litopenaeus vannamei* shrimp – and the bacteria *Vibrio penaeicidan* are promising candidates for the development of a bacterial challenge model. An established breeding line of shrimp, such as the SPF Kona line, ensures little or no genetic variation from one experiment to the next. Additionally, the Kona shrimp are commercially available, thus facilitating the reproducibility of experiments.

V. penaeicida are highly pathogenic to penaeid shrimp. They cause severe infection, produce distinctive histological lesions and are relatively easy to cultivate in vitro. The pathogenicity of *V. penaeicida* has also been studied in *L. stylirostris, Penaeus monodon* and *Marsupenaeus japonicus.*

Virulence study

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The authors recently conducted a series of three consecutive experiments on virulence assessment where juvenile SPF *L. vannamei* from the Kona line were challenged by intramuscular injection with varied concentrations of *V. penaeicida.*

In the first experiment, four groups of 20, 4-gram shrimp received approximately 500,000; 5,000; 500; and 50 colonyforming units (colony-forming unit) bacteria per shrimp, respectively. In the second experiment, four groups of 20 shrimp received approximately 10,000; 5,000; 2,500; and 500 colony-forming unit per shrimp. In the third experiment, four groups of shrimp received approximately 1,000; 500; 250; and 125 colony-forming unit per shrimp.

The two-layer total plate count method was used to estimate bacteria numbers in the concentrated cultures and inocula. A negative control that consisted of 20 shrimp injected with the same sterile 2.5 percent saline used to prepare the inocula was included in each experiment.

The shrimp were maintained in 90-liter tanks at 25-ppt salinity and 27 to 28 degrees-C. Each tank was fitted with its own biological filter. Injected shrimp were monitored every six hours for a maximum of 10 days. Dead shrimp were counted, weighed and discarded. Moribund shrimp were fixed and examined by conventional histology and embryology techniques to confirm the presence of lesions diagnostic of systemic bacterial infection by *V. penaeicida*.

Results

A direct relationship was observed between the bacterial concentrations and shrimp mortality (Fig. 1). At high concentrations, the mortalities were 100 percent for 500,000 colony-forming unit per shrimp, 95 percent for 10,000 colony-forming unit per shrimp, and 90 percent and 91 percent for the two experiments that included 5,000 colony-forming unit per shrimp.



Fig. 1: Mortality of SPF *Litopenaeus vannamei* challenged by intramuscular injection with *Vibrio penaeicida* in three experiments.

At lower concentrations, the mortalities were 95 percent for 2,500 colony-forming unit per shrimp, 25 percent for 1,000 colony-forming unit per shrimp and 25, 20 and 68 percent for the three experiments that included 500 colony-forming unit per shrimp. Shrimp injected with 250 and 125 colony-forming unit per shrimp showed 5 percent mortality, while those at 50 colony-forming unit per shrimp had no mortality.

Similar percentage mortalities have been reported in *L. stylirostris* at concentrations within the 200 to 1,000 colony-forming unit per shrimp range.

The experiments indicated that, while causing significant disease, bacterial concentrations of 500 to 5,000 colonyforming unit per shrimp could allow the shrimp to survive long enough for additional testing. Juvenile Kona *L. vannamei* appear to be an adequate candidate for a challenge model due to their high susceptibility to infection with *V. penaeicida.*

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