



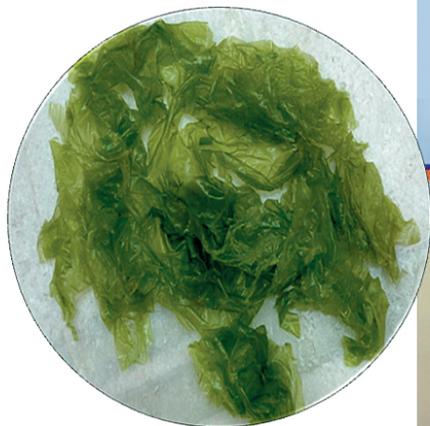
[ANIMAL HEALTH & WELFARE \(/ADVOCATE/CATEGORY/ANIMAL-HEALTH-WELFARE\)](#)

# Single-cell detritus: fermented, bio-enriched feed for marine larvae

Friday, 1 July 2011

By Dr. S. Felix and P. Pradeepa

## Authors' lab is working toward a formulation for black tiger shrimp larvae



Seaweed processed into single-cell detritus is a high-protein product that could supplement or replace microalgae in shrimp hatcheries.

Single-cell detritus (SCD) is a seaweed-based bioproduct produced through a combination of enzymatic and fermentative techniques. It can be prepared at particle sizes of 5 to 12  $\mu$ , making SCD ideal for marine hatchery feeding apart from its bioremediatory and probiotic roles in culture systems.

SCD can be fed to the larvae of both finfish and shellfish. The use of SCD as feed has been studied with oysters and *Artemia* by Motoharu Uchida in Japan. At present, the authors' lab is working toward the formulation and production of SCD for *Penaeus monodon* shrimp larvae as a replacement for unicellular algae.

Trials have so far been successful. The expected breakthrough would be a major development in shrimp hatchery feeding technology by making nutrition management simpler and more cost-effective.

## SCD features

Some of the features that SCD offers include the fact that with crude protein levels up to 35 percent, it is relatively nutritious and could partially or fully replace microalgae as a feed in hatcheries.

SCD particles can be produced in various sizes, as per need and species. The high cell concentration of SCD is comparable to that of algal concentrates. It can act as a bioremediatory agent and has proven probiotic effects.

Mass preparation of SCD is somewhat easier than the production and maintenance of microalgae. The production and use of SCD is an economically viable technology. In addition, single-cell detritus can be stored up to a year at room temperature.

## Fermentation

Fermentation is one of the oldest biotechnological techniques that can be used for marine larval feed preparation. Uchida of Japan was the initiator of the formulation and production of SCD for marine oyster hatcheries.

For maximum utilization of the dietary potential of macroalgae, it is advantageous to perform thalli degradation under conditions regulating the catabolic losses. Mechanical or enzymatic fragmentation is effective for this purpose. Using viable bacteria for degradation is another alternative.

Another interesting characteristic of the detritus diet is the attachment of bacteria to the surface of the detritus, which can be achieved by incubating the bacteria for several hours with axenically prepared SCD particles. This method has some useful functions, such as anti-pathogenic activity and a vitamin-producing ability, and is expected to be useful in the development of a functional hatchery diet for suspension-feeding animals. The combination of lactic acid bacteria and yeast might have a synergistic effect for reducing the prevalence of pathogenic microbes in the production process.

Cellulase enzyme was used initially for the production of single cell units. Fermentation of seaweed was carried out by inoculating a lactic acid bacterium and yeast at a rate of 10<sup>4</sup> cfu/mL. A sugar substrate and nitrogen substrate were added to enhance the rate of fermentation and protein concentration. The process of fermentation was monitored continuously by estimating the lactic acid concentration, and measuring pH, microbial propagation rate and odor.

## Two-phase protocol

SCD production has two phases. The first phase is cellulo-lytic enzymatic treatment of seaweed, which leads to single cell units. The enzymatic digest is further treated with bacteria and yeast in the second, fermentative phase. These two phases can be performed simultaneously or one by one.

### Cellulolytic phase

Algae have cellulose in their cell walls that keeps the cells intact. When cellulose is digested, the individual cells are released and become single cell units. The enzyme cellulase is used for this purpose with the end product of cellulolytic digestion sugar. This phase has two roles: to produce single cell units and to produce sugars for the fermentative phase.

### Fermentative phase

Two organisms are used in the fermentative phase to produce SCD: lactic acid bacteria and yeast. These organisms can be isolated from the natural fermented seaweed or other sources. In 2004, Motoharu Uchida used *Lactobacillus brevis* bacteria, and the yeasts *Debaryomyces hanseii* var. *hanseii* and *Candida zeylanoides* isolated from fermented *Ulva*. Bacteria like *L. plantarum* and *L. casei* can also be used for this purpose. Any suitable source of yeast can be used for fermenting the seaweed.

In the authors' work, a consortium of microbes including *L. plantarum* and *S. cerevisiae* was used to produce 5- to 12- $\mu$  SCD to feed shrimp in a larval-rearing system. Sugar substrate was added to increase the fermentation rate, and a nitrogen source was added to increase the protein concentration, which is essential for shrimp larvae.

Lactic acid bacteria and yeast utilize the sugar produced by cellulolytic digestion and produce lactic acid. This prevents other organisms from growing and thus preserves the SCD.

Lactic acid bacteria also act as a probiotic and thus help to increase survival and maintain water quality. Yeast predominately acts as a bioremediatory agent, which enables culture systems to be run with little or no water exchange.

## Large-scale production

SCD can be produced in large quantities in simple air-tight containers or more sophisticated fermentors or bioreactors specially designed for the purpose. The main difference between the two technologies is that it takes nearly two weeks for SCD to ferment in air-tight containers but only two to three days in a fermentor. Further, for purity and quality production, fermentors are recommended.

*(Editor's Note: This article was originally published in the July/August 2011 print edition of the Global Aquaculture Advocate.)*

## Authors

---



### DR. S. FELIX

Fisheries Research and Extension Centre  
Tamil Nadu Veterinary and Animal Sciences University  
Chennai 600051  
Tamil Nadu, India  
[sugafelix@yahoo.com](mailto:sugafelix@yahoo.com) (mailto:sugafelix@yahoo.com)



**P. PRADEEPA**

Fisheries Research and Extension Centre  
Tamil Nadu Veterinary and Animal Sciences University  
Chennai 600051  
Tamil Nadu, India

Copyright © 2016–2018  
Global Aquaculture Alliance